# Debugging and Modelling of Genetic Circuits

#### **UE2.1 Biological Parts and Devices**



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## How do circuits break?

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(Brophy & Voigt, 2014. Nat. Methods)

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(Brophy & Voigt, 2014. Nat. Methods)

#### **External control over components?**



#### **Diversity and Redundancy in Design?**



• RBS (or promoter) libraries with several expression levels

### **Directly visualize the components?**

• Visualise transcribed RNA directly: malachite green stabilization by RNA aptamer





Aptamer binding

• Visualise translated protein: YFP measurement

(Yerramilli & Kim, 2018. ACS Synbio)

#### **Circuit failures: Under the hood**

• CELLO: Increasing number of parts increases chances of errors



- Fluorescent protein fusions can be used to analyse each step separately
- From this analysis, most of the circuit failures point to unexpected behavior from the anhydrotetracycline (aTc) sensor (seven circuits) or AmtR gate (two circuits).

• Direct analysis of transcription states by RNAseq





• Transcriptional profiling revealed a number of unexpected reasons of failure:

(1) cryptic antisense promoters

(2) terminator failure

(3) sensor malfunction due to media-induced changes in host gene expression



• Translation profiling reveals ribosome occupancy



- Internal start codon can affect genetic circuits
- Eliminating internal translation start site can fix the problem

## What is a model?

#### What is a model?

- Some informative/ conceptual representation of a system
- One or more equations that describe the relationship between different components/ properties of the system
- For example, the Exponential Growth Model:

 $N_{t2} = N_{t1}^{*}(2^{g})$  $\mu = \log (N_{t2} / N_{t1}) / (t2 - t1)$ 



[r = µ]

#### **Growth model: boundless or bounded**

- The Exponential Growth Model assumes infinite resources.
- The Logistic Growth Model defines a **fixed carrying capacity**.



https://www.khanacademy.org/science/ap-biology/ecology-ap/population-ecology-ap/a/exponential-logistic-growth

#### **Biological Modelling**



(Protein Biochemistry & Bioinformatics Dynamic modeling. Stefan Legewie & Sofya Lipnitskaya) https://cbdm.uni-mainz.de/mb18/

## Setting up your computer for simple modelling

- Essential: python
- Useful: jupyter Notebook (local) / Google Collab (cloud)

Some installation / help links:

Introduction to Python

https://biocircuits.github.io/appendices/appB\_python/index.html

Python + Jupyter on Linux WSL in Windows
 <u>https://harshityadav95.medium.com/jupyter-notebook-in-windows-subsystem-for-linux-wsl-8b46fdf0a536</u>

Jupyter Notebook Complete Beginner Guide 2023
 <u>https://www.youtube.com/watch?v=5pf0\_bpNbkw</u>

 Python code in your Google Drive / Colaboratory <u>https://www.freecodecamp.org/news/google-colaboratory-python-code-in-your-google-drive/</u>





https://tinyurl.com/mssb24colab

### **Expression Modelling**

• A system of Ordinary Differential Equations (ODEs) to model gene expression





$\alpha - \alpha + \alpha$	
$I - I$ dilution $\pm I$ degradati	on
i	

Production ( $\beta$ ) and Degradation ( $\gamma$ ) rates

• Some assumptions: well mixed, real concentrations of molecules, first-order decay

### **Expression Modelling**

• A system of Ordinary Differential Equations (ODEs) to model gene expression



$$rac{dx}{dt}=eta-\gamma x$$

γ=	= $\gamma_{\rm dilution}$	$_{1}+\gamma_{ m degradation}$

Production ( $\beta$ ) and Degradation ( $\gamma$ ) rates

• Some assumptions: well mixed, real concentrations of molecules, first-order decay

$$egin{aligned} rac{dx}{dt} &= eta - \gamma x = 0 \ \ \Rightarrow x_{
m ss} &= eta / \gamma \end{aligned}$$

 Steady-state concentration can be determined as a ratio of production and degradation rates

Elowitz: <u>https://biocircuits.github.io/chapters/01\_intro\_to\_circuit\_design.html</u>

#### **ChatGPT: your coding / learning assistant?**



#### **ChatGPT: your coding / learning assistant?**

python	Ů	Copy c
import numpy as no		
import matplotlib pyplot as plt		
import matprotrib.pyprot as pre		
# Define the ODE parameters		
beta = $0.2 \# \beta$		
$gamma = 0.1 \# \gamma$		
# Define the ODE function		
<pre>def ode(x, t):</pre>		
dxdt = beta - gamma * x		
return dxdt		
# Initial condition		
x0 = 0.0		
# Time coop		
# rime span t = np lingpace(0, 10, 100), $\#$ You can adjust the time span as	0000	lad
t - hp. mspace(0, 10, 100) # fou can august the time span as	need	ieu
# Solve the ODE using Fuler's method		
x = np.zeros(len(t))		
x[0] = x0		
<pre>for i in range(1, len(t)):</pre>		
$delta_t = t[i] - t[i-1]$		
$x[i] = x[i-1] + ode(x[i-1], t[i-1]) * delta_t$		
# Plot the solution		
<pre>plt.figure()</pre>		
<pre>plt.plot(t, x)</pre>		
<pre>plt.xlabel('Time (t)')</pre>		
plt.ylabel('x(t)')		

plt.title('Solution of  $dx/dt = \beta - \gamma x'$ )

plt.grid(True)
plt.show()

$$rac{dx}{dt} = eta - \gamma x$$

In this code, we first define the ODE function 'ode(x, t)' as 'dx/dt =  $\beta$  -  $\gamma x$ '. Then, we use Euler's method to numerically solve the ODE over a specified time span. You can adjust the values of 'beta', 'gamma', the initial condition 'x0', and the time span 't' to suit your specific problem. The code also includes a plot of the solution.

#### ChatGPT + Google Colab



#### **Transcription-Translation Modelling**

 A system of Ordinary Differential Equations (ODEs) to model transcription and translation steps



$$egin{aligned} m_{
m ss} &= eta_m/\gamma_m \ x_{
m ss} &= rac{eta_p m_{
m ss}}{\gamma_p} = rac{eta_peta_m}{\gamma_p\gamma_m} \end{aligned}$$

Steady-state values

- Steady-state concentration can be determined as a ratio of production and degradation rates
- Transcription proportionally changes both mRNA and protein levels

#### **Transcription-Translation Modelling**





• Time course of mRNA and protein expression

$$m(t) = m_{ss} \cdot (1 - e^{-\gamma_m t})$$
$$x(t) = x_{ss} \cdot (1 - \frac{\gamma_m e^{-\gamma_p t} - \gamma_p e^{-\gamma_m t}}{\gamma_m - \gamma_p})$$

Instantaneous concentrations

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Elowitz: <u>https://biocircuits.github.io/chapters/01\_intro\_to\_circuit\_design.html</u> Protein Biochemistry & Bioinformatics Dynamic modeling. Stefan Legewie & Sofya Lipnitskaya: <u>https://cbdm.uni-mainz.de/mb18/</u>

#### **Transcription-Translation Modelling**





• Time course of mRNA and protein expression

mRNA degradation rate (γ<sub>m</sub>) changes final steady state and response time

$$m(t) = m_{ss} \cdot (1 - e^{-\gamma_m t})$$
$$x(t) = x_{ss} \cdot (1 - \frac{\gamma_m e^{-\gamma_p t} - \gamma_p e^{-\gamma_m t}}{\gamma_m - \gamma_p})$$

Instantaneous concentrations

mRNA and protein synthesis rates (
$$\beta_m$$
,  $\beta_p$ )  
change only final steady state

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protein degradation rate (γ<sub>p</sub>) changes final steady state and response time

#### **Regulated Transcription: Repressor**



$$K_{
m d}=rac{k_-}{k_+}$$

(Dissociation constant)

$$eta(r)=eta_0rac{p}{p_{ ext{tot}}}=rac{eta_0}{1+r/K_{ ext{d}}}$$



- $\beta_0$  is the unregulated transcription rate
- Assumes separation of time-scales

#### **Regulated Transcription: Repressor**



$$eta(R) = lpha_0 + eta_0 rac{p}{p_{ ext{tot}}} = lpha_0 + rac{eta_0}{1+r/K_{ ext{d}}}$$

(with leaky transcription)



Elowitz: <u>https://biocircuits.github.io/chapters/01\_intro\_to\_circuit\_design.html</u>

#### **Regulated Transcription: Repressor**



$$eta(r)=rac{eta_0}{1+(r/K_{
m d}})^{
m n}$$

(Hill function kinetics)



Elowitz: <u>https://biocircuits.github.io/chapters/01\_intro\_to\_circuit\_design.html</u>

#### **Regulated Transcription: Activator**



$$eta(a) = eta_0 rac{p_{ ext{bound}}}{p_{ ext{tot}}} = eta_0 \, rac{a/K_{ ext{d}}}{1+a/K_{ ext{d}}}$$

(Activated expression)



$$eta(r)=eta_0rac{p}{p_{
m tot}}=rac{eta_0}{1+r/K_{
m d}}$$

(Repressed expression)

- $\beta_0$  is the unregulated max transcription rate
- Assumes separation of time-scales

#### **Regulated Transcription: Activator**



$$eta(a) = eta_0 \, rac{(a/K_{
m d})^{
m n}}{1 + (a/K_{
m d})^{
m n}}$$

(Hill function kinetics)



Elowitz: <u>https://biocircuits.github.io/chapters/01\_intro\_to\_circuit\_design.html</u>

#### **Network Motifs modify circuit behaviour**





NFL speeds up response time



- NFL results in steeper rise in the concentration of protein X.
- NFL reduces noise

#### NFL speeds up response time

$$m(t) = m_{ss} \cdot (1 - e^{-\gamma_m t})$$
$$x(t) = x_{ss} \cdot (1 - \frac{\gamma_m e^{-\gamma_p t} - \gamma_p e^{-\gamma_m t}}{\gamma_m - \gamma_p})$$

Instantaneous concentrations



• NFL results in steeper rise in the concentration of protein X.

#### **Network Motifs modify circuit behaviour**





PFL slows down response time



- PFL results in slower rise in the concentration of protein X.
- PFL increases noise

#### **Transcription-Translation Modelling with Reaction Mechanism**

- In the previous examples the  $\beta_m$ ,  $\beta_x$ ,  $\gamma_m$ ,  $\gamma_x$  are experimentally observed
- Accounting for DNA concentration (CopyN) and leaky transcriptions
- Model-calculated translation rate  $(\beta_x)$



#### ODEs

$$\frac{d[mRNA_{T7RNAP}]}{dt} = CopyN. (primingR_{T7RNAP} + R_{T7prT7RNAP}) - \delta_{mRNA}. [mRNA_{T7RNAP}]$$
(1)  

$$\frac{d[T7RNAP]}{dt} = PR_{T7RNAP}.[mRNA_{T7RNAP}] - \delta_{T7RNAP}. [T7RNAP]$$
(2)  

$$\frac{d[mRNA_{GFP}]}{dt} = CopyN. (leakyR_{GFP} + R_{T7prGFP}) - \delta_{mRNA}. [mRNA_{GFP}]$$
(3)

$$\frac{d[GFP]}{dt} = PR_{GFP} [mRNA_{GFP}] - \delta_{GFP} . [GFP]$$
(4)

(Kushwaha & Salis, 2015. Nat. Comm.)

#### **Expression Modelling: Repressilator**







$$\frac{dx_3}{dt} = \frac{\beta}{1 + (x_2/k)^n} - \gamma x_3$$



β = 10 n = 3

Elowitz: https://biocircuits.github.io/chapters/09 repressilator.html

#### **Expression Modelling: Repressilator**











Elowitz: https://biocircuits.github.io/chapters/09 repressilator.html



#### **TinkerCell**

• A visual modelling tool for computer aided design of genetic circuits <a href="http://tinker-cell.blogspot.com/">http://tinker-cell.blogspot.com/</a>



http://www.tinkercell.com/ (Chandran *et al.*, 2009. J Biol Eng)



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http://www.tinkercell.com/ (Chandran *et al.*, 2009. J Biol Eng)

#### **How many Fitted Parameters?**

• What function will fit this data?



#### **How many Fitted Parameters?**

• Fitting too many parameters may result in less useful models (R<sup>2</sup> notwithstanding)



#### **How many Fitted Parameters?**

• Fitting too many parameters can result in less useful models

"with four parameters I can fit an elephant, and with five I can make him wiggle his trunk." -John von Neumann

(as narrated by Enrico Fermi in Dyson 2004, Nature v427, p297)



(Mayer *et al.*, 2010. American Journal of Physics v78, p648)

#### How detailed does a model have to be?

• A model may be useful even if it does not represent all the details of the system

#### "All models are wrong, but some are useful."

-George Box (1979), "Robustness in the strategy of scientific model building"

"Now it would be very remarkable if any system existing in the real world could be exactly represented by any simple model. However, cunningly chosen **parsimonious** models often do provide remarkably useful approximations. For example, the law PV = RT relating pressure P, volume V and temperature T of an "ideal" gas via a constant R is not exactly true for any real gas, but it frequently provides a useful approximation and furthermore its structure is informative since it springs from a physical view of the behavior of gas molecules. For such a model there is no need to ask the question "Is the model true?". If "truth" is to be the "whole truth" the answer must be "No". The only question of interest is "Is the model illuminating and useful?".

#### **Circuit Modelling and Finding Parameters from Literature**

#### Resources

Biological Circuit Design <a href="https://biocircuits.github.io/index.html">https://biocircuits.github.io/index.html</a>



BioModels Parameters: <u>https://www.ebi.ac.uk/biomodels/parameterSearch</u> (Glont et al., 2020. Bioinformatics.)



• <u>https://bionumbers.hms.harvard.edu/search.aspx</u>

# **Questions welcome.**

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